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SYNTHESIS OF PHOSPHOROTHIOATE ANALOGS OF PHOSPHATIDYLINOSITOL 3,4,5-TRISPHOSPHATE

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Abstract: Phosphorothioate analogs of phosphatidylinositol 3,4,5-trisphosphate have been synthesized using Me₃N-Me₃SiCl reagent in a critical exhaustive cleavage of *O*-benzyl phosphorothioate triester intermediates. © 1997 Elsevier Science Ltd.

Phosphatidylinositol phosphates (PIP) are important precursors of inositol phosphate second messengers, or are the cellular signals themselves.¹ In particular, the 3-phosphorylated phosphatidylinositols **1-3** have been implicated in interactions with various cellular targets to mediate diverse physiological events ranging from vesicular trafficking to insulin response.² The primary metabolic pathways of the 3-phosphorylated PIs involve a series of specific phosphatases rapidly removing phosphates from the 5-, 4- and 3-positions to finally form phosphatidylinositol.^{2,3} One approach to study the function of these phospholipids would be to increase their biological lifetime, by making them resistant to specific phosphatases or phosphodiesterases. Such resistance can be conferred by substituting the phosphate groups with the corresponding phosphorothioates. Phosphorothioate analogs of phosphate monoesters and diesters have been long known to undergo slower enzymatic hydrolysis than their phosphate counterparts,⁴ and the phosphatase-resistant phosphorothioate analogs of inositol 1,4,5-trisphosphate (IP₃) have been previously used to investigate the role of IP₃ in calcium release.⁵ This work describes the first synthesis of phosphorothioate analogs of phosphates **4** and **5**.⁶

$$\begin{array}{c} R_{4}COO \\ R_{4}COO \\ \\ R_{1} = PO_{3}^{-2}, \ R_{2} = R_{3} = H, \ X = O, \ PI-3-P, \ 1 \\ R_{1} = R_{2} = PO_{3}^{-2}, \ R_{3} = H, \ X = O, \ PI-3,4-P_{2}, \ 2 \\ R_{1} = R_{2} = R_{3} = PO_{3}^{-2}, \ X = O, \ PI-3,4-S-P_{3}, \ 3 \\ R_{4} = alkenyl, \ alkyl \\ R_{1} = R_{2} = R_{3} = PO_{2}S^{-2}, \ X = O, \ PI-3,4-S-P_{3}, \ 4 \\ R_{1} = R_{2} = R_{3} = PO_{2}S^{-2}, \ X = S, \ PSI-3,4-S-P_{3}, \ 5 \\ R_{4} = C_{15}H_{31} \end{array}$$

Despite the close structural analogy between phosphates and phosphorothioates, synthesis of the latter is more difficult. For example, a convenient hydrogenolytic deprotection of phosphate groups used by all methods reported to date⁶ is not applicable in the presence of sulfur, and the instability of the P-S bond makes also a number of other deprotection methods unsuitable. Below we present a deprotection scheme that enables synthesis of phosphorothioate monoesters in the polyphosphate environment, in the presence of the labile acyl ester groups.

The previously synthesized triol $6^{\text{vd},7}$ was treated with O,O-dibenzyl-N,N-diisopropylphosphoramidite to give the corresponding 3,4,5-trisphosphite 7. This compound was added with elemental sulfur to produce the trisphosphorothioate 8, which was consecutively treated with tetra-n-butylammonium fluoride to afford the

alcohol **9**. The reaction of this alcohol with *N*,*N*-diisopropyl-*O*-methyl phosphoroamidochloridite, followed by condensation of the resulting phosphoramidite with 1,2-dipalmitoyl-*sn*-glycerol (DPG) and oxidation with dinitrogen tetroxide or sulfurization gave the fully protected PI-3,4,5-Ps₃ **10** and the fully protected PsI-3,4,5-Ps₃ **11**, respectively.

(i) $iPr_2N-P(OBn)_2$, tetrazole; (ii) S_8 ; (iii) $Bu_4N^+F^-$; (iv) $Cl-P(OMe)(NiPr_2)$, $EtiPr_2N$; (v) DPG, tetrazole; (vi) N_2O_4 ; vii: S_8 ; (viii) Me_3N

The demethylation of the phosphate group in the 1-position was performed using trimethylamine to give the corresponding phosphodiester group. This reagent was found to also simultaneously cleave benzyl groups from the *O,O*-dibenzylphosphorothioate residues at the 3, 4- and 5-positions to give the corresponding 3,4,5-tris-*O*-benzylphosphorothioates **12** and **13**. Due to the presence of chiral centers at each phosphorothioate group these products were mixtures of stereoisomers giving rise to complex ³¹P NMR spectra. Further dealkylation of a phosphodiester to a phosphomonoester does not occur with this (and other nucleophilic reagents) due to a poor leaving group character of a phosphorothioate (phosphate) monoester dianion; a further conversion to a monoester requires some protic or Lewis acid catalysis.

The typical approaches to cleave the mono-O-benzyl phosphorothioate group proved problematic. First, the debenzylation of the phosphodiester groups in 12 with ethanethiol-BF₃ was unsuccessful. The reaction with this reagent yielded large amounts of by-products (50%) giving rise to the ³¹P NMR signals at 20-30 ppm, and additional by-products at ca. -10 ppm. The range of the observed chemical shifts suggested that in addition to the O-benzyl ester cleavage, a partial thiono-thiolo rearrangement⁸ took place to give the corresponding S-benzyl derivatives 14 (δ ca. 20 ppm). These thiolesters could then undergo a nucleophilic displacement of the S-benzyl group by the adjacent phosphorothioate nucleophile to give a cyclic pyrophosphate 15 (δ ca. -10 ppm, see the scheme below). The attempted solvolysis of benzyl esters in TFA/CHCl₃ produced only the rearrangement and cyclization products 14 and 15. The reaction of the phosphorothioate 12 with TMS iodide resulted in a partial

cleavage of the phosphodiester bond at the 1-position, rendering this potential method of deprotection unsuitable, as well. It has to be stressed, that due to the fact that all three phosphorothioate groups have to be deprotected to give the desired product, but that isomerization at any single phosphorothioate function produces an unusable side-product, a relatively low rate of isomerization translates into large losses of yield. Furthermore, the very polar nature of the final products and their low solubility in organic media essentially preclude product purification by typical chromatographic methods.

In contrast, the treatment of both esters 10 and 11 with the mixture of trimethylamine-TMS chloride afforded exclusively the desired persilylated derivatives of tris-monophosphorothioates 16 and 17, respectively. The rationale for using this reagent is to persilylate the initially formed tris-O-benzyl diester 12 to form the neutral tris-O-benzyl-O-trimethylsilylphosphorothioate 18. This triester underwent further debenzylation, albeit at a slower rate than the original dibenzyl phosphate. The reaction was followed by ³¹P NMR and the progress of dealkylations at the 3,4,5-positions and at the 1-position could be precisely evaluated due to the 10-13 ppm upfield shift upon replacement of each alkyl group by the silyl group, and due to the fact that only silylation at the oxygen atom (not sulfur) was observed. Thus, the removal of the first benzyl group resulted in the shift from ca. 70 ppm (O,O-dibenzyl phosphorothioate group in 10) to 57 ppm (O-TMS-O-benzyl phosphorothioate group in 18), and the removal of the second group gave a signal at 44 ppm (O,O-bisTMSphosphorothioate group in 16). Analogously, the removal of the methyl group from the phosphate at the 1-position in the derivative 10 resulted in the shift from 2 ppm to -8 ppm. Since these dealkylations were neither regio- nor stereo-selective, a large number of isomeric compounds was formed at the intermediate reaction times, which eventually converged into two diastereomeric species of the product, due to a random silylation of the pro-R and pro-S oxygen atoms of the phosphate group at the 1-position in the product 16.

i: TMS-Cl/Me₃N (1:4 w/w, used as solvent), 23°C, 10 days; ii: acetate buffer; iii: EtSH/BF₃

The cleavage of the O-silyl derivatives 16 and 17 was achieved by aqueous hydrolysis at neutral pH to give exclusively MOM-ethers 20 and 21. It is important that the neutral pH be maintained during the hydrolysis, as

the hydrolysis under unbuffered conditions lead to desulfurization and formation of phosphoanhydrides, apparently by the displacement of the sulfhydryl group by the adjacent phosphorothioate group, analogously as shown above. The MOM-derivatives **20** and **21** were finally deprotected by ethanethiol-BF₃ at room temperature to give the final products **4** and **5**, respectively. The obtained compounds **4** and **5** were fully characterized by ¹H and ³¹P NMR and ES MS.^{10,11} Their evaluation as potential inhibitors of inositol-related enzymes and metabolically stable analogs of PIP₃ is currently underway. We believe that in view of the mild character of the deprotection conditions described here and the lability of the synthesized analogs of PIP₃, this scheme should be well applicable to synthesis of other phosphorothioate monoester analogs of complex natural phosphates.

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- 11. The 1;1 ratio of diastereomers of the product 5 is indicated by the ¹H NMR spectrum of the derivative 11.
- 12. **4** (NH₄*): ¹H NMR (CDCl₃:CD₃OD:D₂O, 1:1:0.3) δ 0.89 (t, *J* = 6.7 Hz, 6 H), 1.22-1.38 (m, 48 H), 1.52-1.68 (m, 4 H), 2.29-2.37 (m, 4 H), 3.94-3.99 (m, 1 H), 4.04-4.11 (m, 3 H), 4.18-4.24 (m, 1 H), 4.32-4.37 (m, 1 H), 4.43 (m, 1 H), 4.52 (m, 1 H), 4.82 (m, 1 H), 5.27 (m, 1 H) (the two missing inositol protons are most likely overlapped by the broad OH signal); ³P NMR (CDCl₃:CD₃OD:D₂O, 1:1:0.3) δ -0.61 (1P), 52.4 (2P), 53.2 (1P); ES MS [M-H]*=1097. **5** (NH₄*); ³H NMR (CDCl₃:CD₃OD:D₂O, 1:1:0.3) δ 0.89 (t, *J* = 6,6 Hz, 6H), 1.27 (m, 48 H), 1.51-1.68 (m, 4 H), 2.29-2.36 (m, 4 H), 3.93-4.01 (m, 1 H), 4.03-4.38 (m, 6 H), 4.41-4.46 (m, 1 H), 4.79 (m, 1 H), 5.22-5.34 (m, 1 H); the missing inositol proton is most likely overlapped by a broad OH resonance; ³¹P NMR (CDCl₃:CD₃OD:D₂O, 1:1:0.3) δ 52.3, (1P), 52.6 (1P), 53.2 (1P), 56.7 (1P); ES MS [M-H]*=1113.